

# Activity of an Investigational Polymyxin-B-Like Compound (SPR206) against a Set of *Enterobacteriaceae* Organisms Responsible for Human Infections

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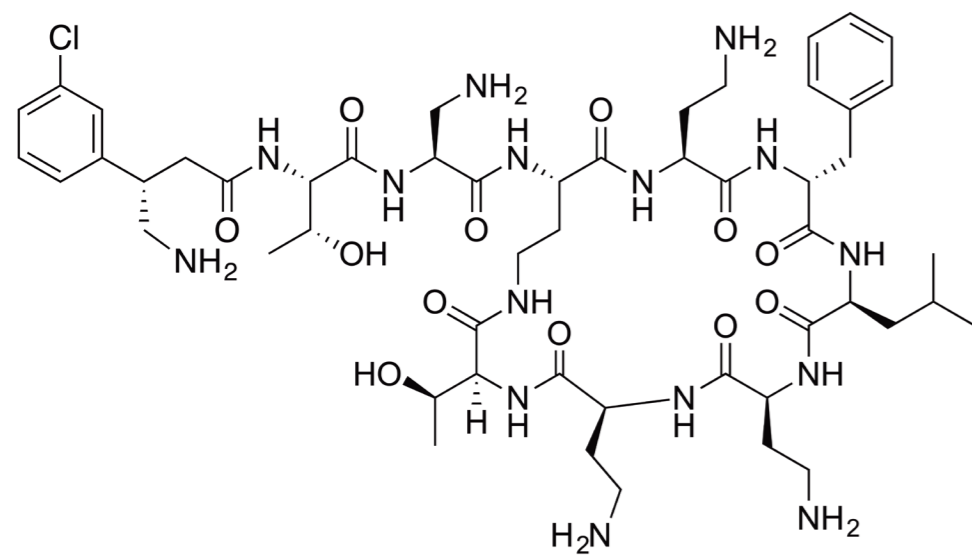


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## Introduction

- Gram-negative bacteria producing extended-spectrum β-lactamase (ESBL) and/or carbapenemase enzymes that show resistance to many antibiotics have been steadily increasing to alarming levels in hospital and community settings
- SPR206 is a next-generation polymyxin compound being developed for treating infections caused by gram-negative organisms (Figure 1)
- This study evaluated the *in vitro* potency of SPR206 and compared its potency to those of polymyxin-B and colistin against *Enterobacteriaceae*, including carbapenem-resistant (CRE) organisms

Figure 1 Structure of SPR206



## Results

- SPR206 (MIC<sub>50/90</sub>: 0.06/0.12 mg/L) was more potent than colistin and polymyxin-B (MIC<sub>50/90</sub>: 0.25/0.25 mg/L; Table 1, Figures 2 and 3)
  - SPR206 inhibited 93.2% of all *Enterobacteriaceae* at ≤0.12 mg/L, while colistin and polymyxin-B inhibited 38.3% and 33.1%, respectively, at ≤0.12 mg/L (Table 1)
- SPR206 had an MIC<sub>100</sub> of ≤2 mg/L against *Escherichia*, *Citrobacter*, *Salmonella*, and *Shigella* species (Table 1)
- Ceftriaxone displayed a bimodal MIC distribution (MIC<sub>50/90</sub>: ≤0.12/>8 mg/L) against all *Enterobacteriaceae* isolates and 77.4% were susceptible at the CLSI and EUCAST breakpoints of ≤1 mg/L
- Meropenem was very active (MIC<sub>50/90</sub>: ≤0.12/≤0.12 mg/L) against these isolates and 97.0%/97.2% were susceptible at the CLSI/EUCAST breakpoints, respectively (Table 1)
- Against a CRE challenge set, SPR206 (MIC<sub>50/90</sub>: 0.06/0.12 mg/L) showed MIC values 4-fold lower than colistin and polymyxin-B (MIC<sub>50/90</sub>: 0.25/0.5 mg/L; Table 1)
  - Isolates included *bla*<sub>KPC</sub>, *bla*<sub>NDM</sub>, *bla*<sub>VIM</sub>, and *bla*<sub>OXA-48</sub> genotypes
  - MIC results similar to the random selection set are seen in Table 1
    - As expected, ceftriaxone (MIC<sub>50/90</sub>: >8/>8 mg/L) and meropenem (MIC<sub>50/90</sub>: >8/>8 mg/L) showed little activity against this challenge set (Table 1)

## References

- Clinical and Laboratory Standards Institute (2018). *M100Ed28E. Performance standards for antimicrobial susceptibility testing: 28th informational supplement*. Wayne, PA: CLSI.
- Clinical and Laboratory Standards Institute (2018). *M07Ed11E. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard—eleventh edition*. Wayne, PA: CLSI.
- EUCAST (2018). Breakpoint tables for interpretation of MICs and zone diameters. Version 8.0, January 2018. Available at [http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Breakpoint\\_tables/v\\_8.0\\_Breakpoint\\_Tables.pdf](http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_8.0_Breakpoint_Tables.pdf). Accessed January 2018.

Figure 2 Comparison of colistin to SPR206 when tested against 573 *Enterobacteriaceae* isolates

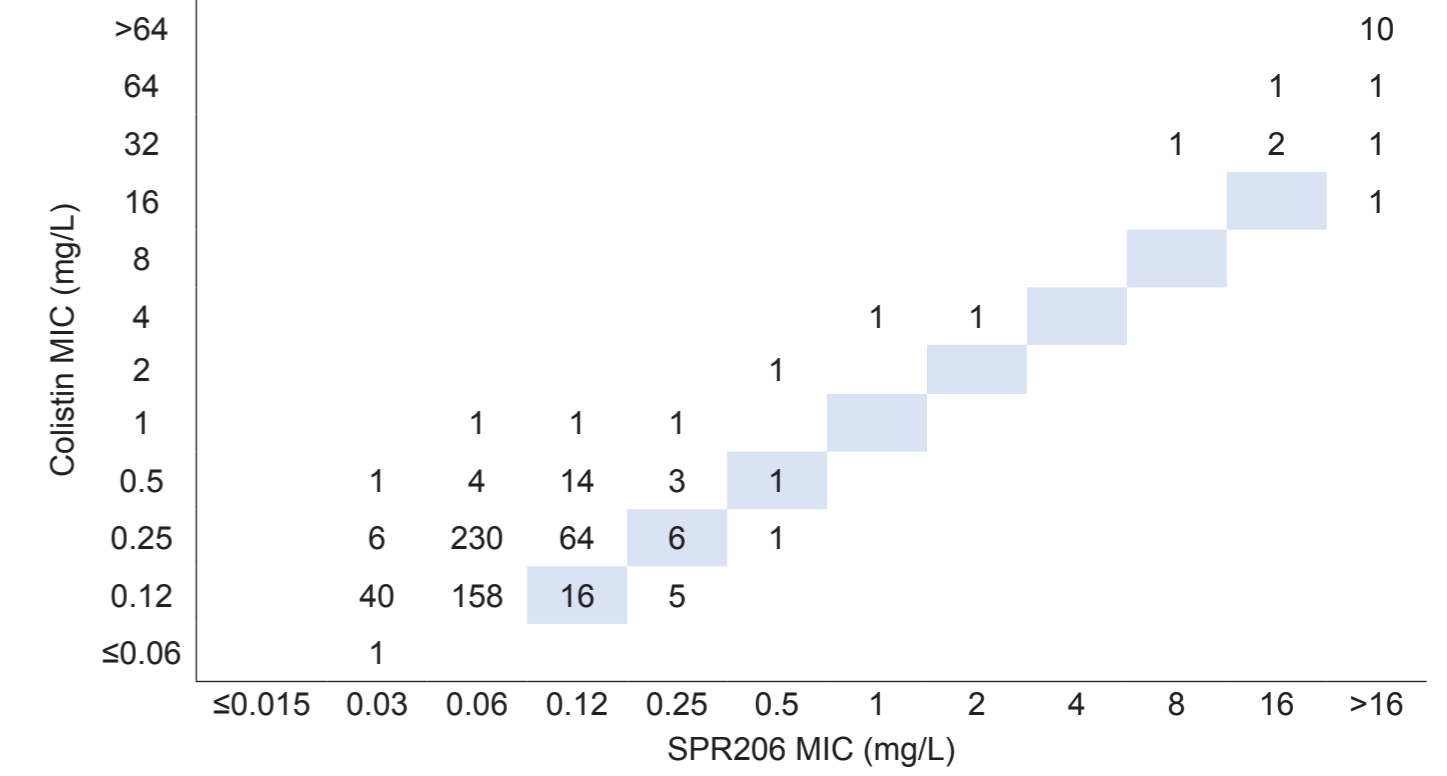
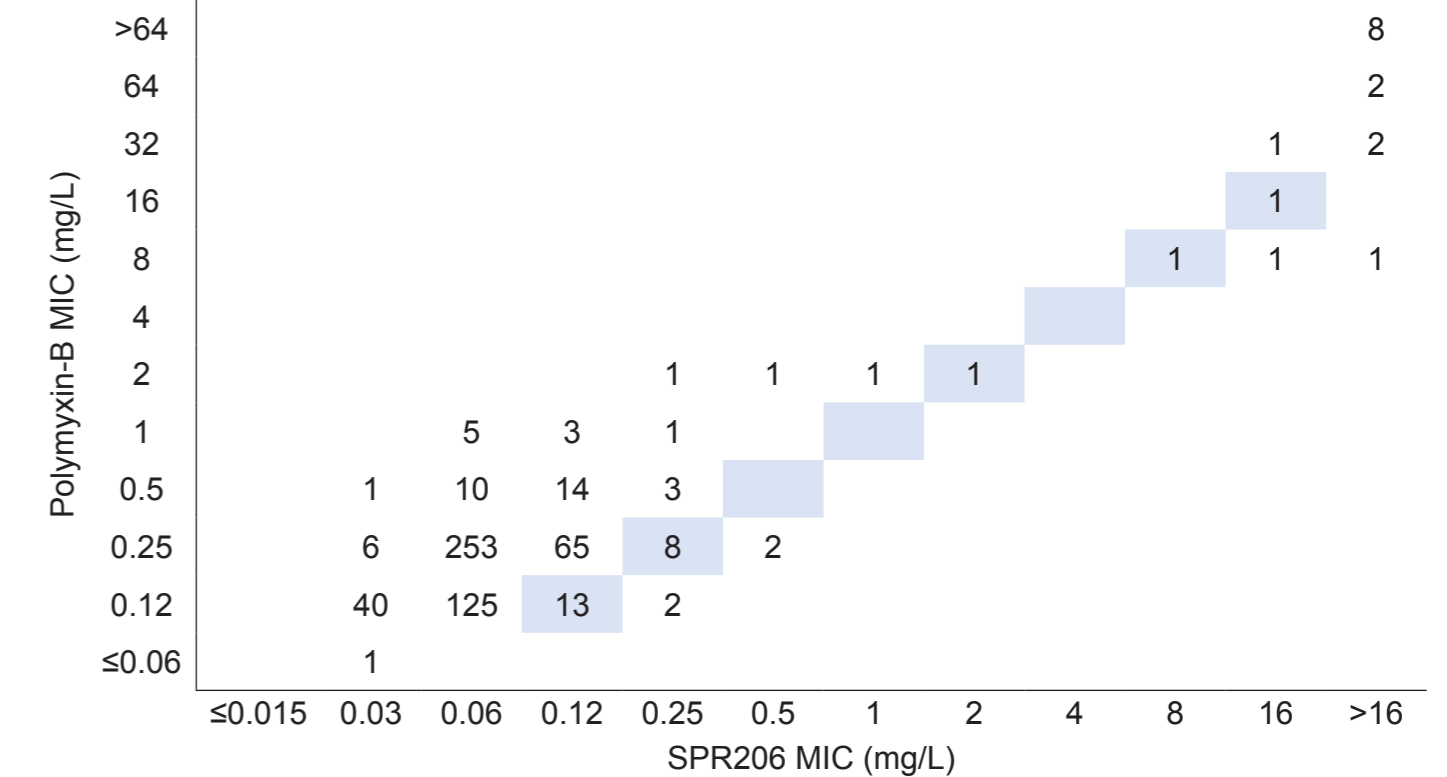


Figure 3 Comparison of polymyxin-B to SPR206 when tested against 573 *Enterobacteriaceae* isolates



## Materials and Methods

### Bacterial isolates

- A total of 541 recent clinical *Enterobacteriaceae* isolates (2016–2017) were randomly selected through the SENTRY Antimicrobial Surveillance Program from 150 medical centers worldwide
- Isolates were responsible for bloodstream (30%), urinary tract (26%), pneumonia (20%), skin and skin structure (15%), and other infections (9%)
- Drug activities were also investigated against an independent challenge set of 32 CRE isolates (Table 1)
- Isolates were determined to be clinically significant based on local guidelines and submitted to a central monitoring laboratory (JMI Laboratories, North Liberty, Iowa)
- Bacterial isolate identification was confirmed by standard algorithms supported by matrix-assisted laser desorption/ionization-time of flight mass spectrometry (Bruker Daltonics, Bremen, Germany)

### Antimicrobial susceptibility testing

- Isolates were tested for susceptibility by broth microdilution following guidelines in the CLSI M07 (2018) document
- Frozen-form reference 96-well panels manufactured by JMI Laboratories were used for testing
- Breakpoint criteria for comparator agents were from the CLSI M100 (2018) and EUCAST (2018) documents

## Conclusions

- Overall, SPR206 was highly potent against a contemporary collection of *Enterobacteriaceae* isolates
- Based on MIC<sub>50/90</sub> results, SPR206 potency was consistently 2- to 4-fold greater than the potency of colistin and polymyxin-B
- Against a challenge set of isolates with increased carbapenem MIC values:
  - SPR206 MIC results were not adversely affected when compared with the MIC values obtained against randomly selected organisms.
  - SPR206 MIC values were consistently lower than colistin and polymyxin-B
- These *in vitro* results obtained for SPR206 warrant its further development as an option for treating gram-negative infections

## Acknowledgements

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Table 1 Antimicrobial activity of SPR206 and comparators tested against the main organisms and organism groups

Organism group (no. of isolates)	No. and cumulative % of isolates at MIC (mg/L) of <sup>a</sup> :														MIC <sub>50</sub>	MIC <sub>90</sub>	EUCAST %S				
	≤0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	> b							
<b>Enterobacteriaceae (541)</b>																					
SPR206	47	8.7	370	77.1	87	93.2	15	95.9	3	96.5	1	96.7	1	96.9	0	97.0	13	100.0	0.06	0.12	
Colistin			1	0.2	206	38.3	293	54.2	18	63.3	3	66.5	1	67.6	2	69.7	4	74.1	0.25	0.25	96.5
Polymyxin-B			1	0.2	178	32.7	310	57.3	25	61.4	6	67.5	4	69.9	0	70.0	3	75.4	0.25	0.25	
Ceftriaxone					348	64.3	42	72.1	21	76.0	8	77.4	3	78.6	3	79.7		110	≤0.12	>8	77.4
Meropenem					516	95.4	6	96.5	3	97.0	0	97.2	0	97.2	0	97.2		14	≤0.12	≤0.12	97.2
<b>E. coli (182)</b>																					
SPR206		15	8.2	134	81.9	30	98.4	1	98.9	0	99.5	0	99.5	1	100.0				0.06	0.12	
Colistin					39	21.4	135	74.2	7	95.6	0	99.5	0	99.5	0	100.0			0.25	0.25	99.5
Polymyxin-B					56	30.8	114	62.6	8	93.4	3	97.8	1	99.5	1	100.0			0.25	0.25	
Ceftriaxone					143	78.6	3	80.2	0	80.2	1	80.8	0	80.8	0	81.9		33	≤0.12	>8	80.8
Meropenem					182	100.0												14	≤0.12	≤0.12	100.0
<b>K. pneumoniae (181)</b>																					
SPR206		9	5.0	146	85.6	19	96.1	4	98.3	0	98.3	0	98.3	0	98.3	2	99.4		0.06	0.12	
Colistin					47	26.5	90	50.3	2	97.8	1	98.3	0	98.3	0	98.3	2	99.4	0.25	0.25	98.3
Polymyxin-B					60	33.1	108	60.2	7	92.8	2	97.8	1	99.5	1	100.0			0.25	0.25	
Ceftriaxone					122	67.4	8	71.8	3	73.5	1	74.0	1	74.6	2	75.7		44	≤0.12	>8	74.0
Meropenem					166	91.7	1	92.3	0	92.3	0	92.8	0	92.8				13	≤0.12	≤0.12	92.8
<b>E. cloacae species complex (94)</b>																					
SPR206		5	5.3	57	66.0	14	80.9	4	85.1	1	86.2	0	86.2	0	86.2	1	88.3		0.06	>16	
Colistin					41	43.6	37	83.0	2	85.1	0	86.2	0	86.2	0	87.2	2	89.4	0.25	0.5	86.2
Polymyxin-B					27	28.7	48	79.8	4	84.0	1	86.2	1	86.2	0	88.3	2	90.4	0.25	0.5	32
Ceftriaxone					20	21.3	19	41.5	5	59.6	2	64.9	3	70.2	1	71.3		27	0.5	>8	64.9
Meropenem					85	90.4	3	94.7	0	97.9	0	97.9	0	97.9	0	98.9		1	≤0.12	≤0.12	97.9
<b>E. aerogenes (22)</b>																					
SPR206			12	54.5	7	86.4	1	90.9	1	95.5	0	95.5	0	95.5	0	95.5		1	0.06	0.25	

<sup>a</sup> The intensity of shading is proportional to the number of tested isolates within each row that display the indicated MIC value.  
<sup>b</sup> Greater than the highest concentration tested.  
<sup>c</sup> Includes *Citrobacter freundii* species complex (4 isolates; 1 *bla*<sub>KPC-2</sub>, 1 *bla*<sub>KPC-3</sub>, 1 *bla*<sub>OXA-48</sub>, and 1 *bla*<sub>VIM-1</sub>), *E. cloacae* species complex (9 isolates; 1 *bla*<sub>KPC-2</sub>, 2 *bla*<sub>KPC-3</sub>, 1 *bla*<sub>KPC-4</sub>, 3 *bla*<sub>NDM-1</sub>, and 1 *bla*<sub>VIM-1</sub>), *Escherichia coli* (9 isolates; 3 *bla*<sub>KPC-2</sub>, 2 *bla*<sub>KPC-3</sub>, 1 *bla*<sub>NDM-1</sub>, 1 *bla*<sub>NDM-5</sub>, and 1 *bla*<sub>OXA-232</sub>), 1 *bla*<sub>NDM-7</sub>, and 1 *bla*<sub>OXA-48</sub>), *Klebsiella oxytoca* (1 isolate, *bla*<sub>KPC-2</sub>), and *K. pneumoniae* (9 isolates; 1 *bla*<sub>KPC-2</sub>, 3 *bla*<sub>KPC-3</sub>, 2 *bla*<sub>NDM-1</sub>, and 3 *bla*<sub>OXA-48</sub>).